

# Determination of Centromere Number of Chromosomes Involved in Robertsonian Translocation by using Fluorescence in Situ Hybridization (FISH)

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## ABSTRACT

Robertsonian translocation is one of the major chromosomal re-arrangements and constitutes 18% of all genetic abnormalities with an incidence of 1/1000 in the general population. Robertsonian translocations are characterized centric fusion of the long arms of the acrocentric chromosomes and generally occur between chromosome 13q, 14q and 14q,21q. In this study, the centromere numbers of 22 patients with Robertsonian translocations were evaluated by using fluorescence in situ hybridization with repetitive DNA probes localized on the centromeric region of acrocentric chromosomes. We detected that all Robertsonian translocation have a single centromere. However some of case showed a distinct size of FISH signals on centromere region, which may indicate the duplication or two different satellite DNA sequences. Therefore these cases should be clarified for the composition of monocentric centromere sequence.

**Key Words:** Robertsonian translocation, centromere, dicentric chromosome, FISH

**Robertsonian Translokasyonlarda İçe Karışan Kromozomların Sentromer Sayısının Floresan in Situ Hibridizasyon (FISH) ile Belirlenmesi**

## ÖZET

Robertsonian translokasyon major kromozomal yeniden düzenlenmelerden biridir. Bu anomali genel popülasyonda 1/1000 sıklık ile tüm genetik anomalilerin % 18'ini oluşturur. Robertsonian translokasyonlar, akrosentrik kromozomların uzun kollarının füzyonu ile karakterize ve genellikle kromozom 13q ve 14q, 14q ve 21q arasında meydana gelmektedir. Mevcut çalışmada 4 farklı tipde 22 Robertsonian translokasyon taşıyıcı ve hasta bireyin, akrosentrik kromozomlarındaki sentromer sayıları bu bölgelere spesifik repetitif DNA problemleri ile floresan in situ hybridization tekniği kullanılarak incelendi. Elde edilen FISH sonuçlarına göre Robertsonian translokasyonların tümünün monosentrik olduğu tesbit edildi. Bununla birlikte vakaların bazılarının sentromer bölgesinde farklı büyüklükte FISH sinyalleri gözlemlendi. Bu farklı yapının sebebi sentromeri oluşturan dizilerin duplikasyonu veya iki farklı satellit DNA sekansı olabilir. Sonuç olarak, çalışmaya dahil olan tüm vakalar monosentrik olmakla birlikte, monosentrik yapıyı oluşturan satellit DNA dizilerinin karakterize edilmesi gerekmektedir.

**Anahtar Kelimeler:** Robertsonian translokasyonlar, sentromer, disentrik kromozom, FISH

## INTRODUCTION

Robertsonian translocations are one of the most common structural chromosomal aberrations observed in humans and occur in approximately 1 in every 1000 newborns (1). This translocation can be either de novo or be transmitted by one of carrier parent. Robertsonian translocations are characterized centric fusion of the long arms of the acrocentric chromosomes 13, 14, 15, 21 and 22, with the translocated chromosome bearing either one or two centromeres and resulting in a 45 or 46 chromosome karyotype. The majority of Robertsonian translocations involve two nonhomologous acrocentric chromosomes. The most frequent Robertsonian translocations are der t(13q;14q) and t(14q;21q) have an estimated frequency of 0.97 and 0.20, respectively (2). The acrocentric chromosomes in man have been intensively investigated, both because of their frequent involvement in chromosomal aberrations and aneuploidy and because of their unique structural properties (3). Rearrangements of the acrocentric chromosomes are associated with an increased risk of aneuploidy. In generally, carriers of Robertsonian translocations are phenotypically normal but are at increased risk for infertility, spontaneous abortions or chromosomally unbalanced offspring. Malsegregation of Robertsonian translocation, results in trisomy or monosomy of complete chromosomes (4).

The centromere regions are mystery for different scientists; for cytologists, it is the primary constriction of the mitotic chromosome, for geneticists it is the origin from which recombination distances are measured. Other equally valid definitions include the following: a specialized DNA sequence, a specialized type of heterochromatin, a structure that regulates sister-chromatid-pairing, a structure that attaches the chromosome to the ends of growing or shrinking microtubules, a marshalling region where chromosomal passenger proteins congregate prior to their function elsewhere during the closing stages of mitosis, and a signaling device that monitors chromosomal alignment and tells the cell when it is safe to segregate sister chromatids in mitosis and homologs in meiosis (5). Molecular studies have shown that the pericentromeric and short-arm regions of acrocentric chromosomes have extensive sequence homology, although some sequences are not common to all of the acrocentrics. Several distinct and tandemly repetitive sequences are localized to these regions. Alpha-satellite DNA sequence is a major class of repetitive DNA found at the centromeric region of each human chromosome (6). Centromeres can

be involved in chromosomal anomalies. One of those is Robertsonian translocations. Robertsonian translocations have either single or two centromeres. Therefore we aimed to evaluate the centromer numbers in the patients having Robertsonian translocation by using fluorescence in-situ hybridization (FISH) techniques with acrocentric centromere-specific probes.

## MATERIAL AND METHODS

### *Patients*

This study includes total 22 unrelated (13 female and 9 male) patients with Robertsonian translocation. Twelve of them were Robertsonian translocation carriers including 3 t(14;21) and 9 t(13;14) who have repeated miscarriages and infertility history. The other 10 patients were Down syndrome. Five of them were t(14;21), 3 t(21;21) and 2 t(15;21). The age of the patients ranged from one month to 28 years.

### *Fluorescence in Situ Hybridization (FISH)*

In this study, patients consulting for different purposes such infertility, repeated miscarriages, and birth of a malformed child were included. Conventional cytogenetic analysis in lymphocyte chromosomes revealed a chromosomal translocation for each patient. Briefly, standard chromosome preparations from blood lymphocytes of the carrier and the control groups were used. In order to verify the number of centromere on Robertsonian translocation, FISH analyses were done by using D13Z1/D21Z1 and D14Z1/D22Z1 and D15Z1 probes (Cambio, UK). D13Z1/D21Z1 was labeled with Cy3 (fluoresces red), and D14Z1/D22Z1 was labeled with biotin and then was detected with avidin conjugated with fluorescein isothiocyanate (FITC; fluoresces green) according to previously published procedures (7). For the samples, the probe mixtures were denatured at 70°C for 10 min, while DNA was denatured in 70%formamide/2 SSC at 70°C for 2-3 min. The slides were then dehydrated through an ascending alcohol series (70, 85, and 100%), and airdried. The denatured probe mixture was applied onto the slides. The hybridization was performed in 40°C moisture chamber overnight. The slides were washed twice for 5 min with 2 SSC, 50% formamide/2xSSC and 2xSSC at 42 °C. They were then stained with a counterstain medium containing DAPI (40,6-diamidino-2-phenyl-indole).

**Table 1.** Karyotypes, circumstances of discovery of structural chromosomal abnormality and centromer constitution used to perform FISH of 22 patient or carriers of Robertsonian translocation.

Patient	Karyotype	Discovery	FISH probe	Centromer constitution
1	46, XX, t (21;21)	Down Sendromu	D13Z1/D21Z1	Monocentric
2	46, XX, t (21;21)	Down Sendromu	D13Z1/D21Z1	Monocentric
3	46, XX, t (21;21)	Down Sendromu	D13Z1/D21Z1	Monocentric
4	46, XY, t (14;21)	Down Sendromu	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
5	46, XX, t (14;21)	Down Sendromu	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
6	46, XX, t (14;21)	Down Sendromu	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
7	46, XY, t (14;21)	Down Sendromu	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
8	46, XY, t (14;21)	Down Sendromu	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
9	46, XY, t (15;21)	Down Sendromu	D13Z1/D21Z1 and D15Z1	Monocentric
10	46, XY, t (15;21)	Down Sendromu	D13Z1/D21Z1 and D15Z1	Monocentric
11	45, XX, t (14;21)	Repeated miscarriages	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
12	45, XX, t (14;21)	Repeated miscarriages	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
13	45, XY, t (14;21)	Repeated miscarriages	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
14	45, XX, t (13;14)	Repeated miscarriages	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
15	45, XX, t (13;14)	Repeated miscarriages	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
16	45, XX, t (13;14)	Repeated miscarriages	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
17	45, XX, t (13;14)	Repeated miscarriages	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
18	45, XX, t (13;14)	Repeated miscarriages	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
19	45, XY, t (13;14)	Primary infertility	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
20	45, XY, t (13;14)	Oligoasthenoteratozoospermia	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
21	45, XY, t (13;14)	Oligoasthenoteratozoospermia	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric
22	45, XY, t (13;14)	Oligoasthenoteratozoospermia	D13Z1/D21Z1andD14Z1/ D22Z1	Monocentric

The slides were examined with an epifluorescence microscope (Nikon, Optiphot) equipped with a set DAPI, fluorescein isothiocyanate (FITC) and rhodamine filters. These filters allow individual detection of red and green.

## RESULTS

In this study, we analysed centromeric number in 22 unrelated (13 female and 9 male) Robertsonian translocation patients using FISH technique. FISH analysis

on metaphase and interphase cells from the patient's lymphocyte using probes to the centromeres of chromosomes 13/21, 14/22 and 15 revealed that the translocation had a hybridization signal from each probe, indicating that these rearrangements are monocentric. The results obtained from both conventional cytogenetic and FISH analysis with chromosome specific probes are given in Table 1. FISH signals on some translocations chromosomes were appeared as in distinct size.

## DISCUSSION

The centromere is an essential functional domain responsible for the correct inheritance of eukaryotic chromosomes during cell division. Molecular studies have shown that the pericentromeric and short-arm regions of these five pairs of acrocentric chromosomes have extensive sequence homology, although some sequences are not common to all of the acrocentrics. Several distinct and tandemly repetitive sequences are localized to these regions. Alpha-satellite is a major class of repetitive DNA found at the centromeric region of each human chromosome (6). Acrocentric chromosomes contain a number of satellite subfamilies, some of which are shared by different acrocentrics. These shared sequences have been postulated to be involved in the formation of Robertsonian translocations and in nonrandom participation of chromosomes 13, 14, and 21 in most Robertsonian translocations (1).

The short arms of acrocentric chromosomes can be divided into three distinct portions. The proximal short arm (p11) contains satellites I-IV (8, 9), beta-satellite (10) and the interspersed 724 repeated sequence (11). The stalk, or nucleolus organizer region (NOR) (p12), contains the 18S and 28S ribosomal genes (12). Finally, beta-satellite DNA and 724 DNA have been mapped to the distal cytological satellite (p13). In Robertsonian translocation, single or double centromeres may be formed depending on the breakpoint of chromosomal regions. These centromeric composition and centromer number have an effect on chromosomal segregation, resulting in the further abnormalities.

In the present study, we analysed centromeric number in 22 unrelated (13 female and 9 male) Robertsonian translocation patients or carriers who have four different type of this by using FISH technique. It has been determined that centromeric number is structurally monocentric in all of Robertsonian translocations. FISH analysis on metaphase cells from the patient's lymphocyte culture using probes to the centromeres of chromosomes 13, 14, 15, 21 and 22 revealed that Robertsonian translocations had a hybridization signal from each probe, indicating that this rearrangements is monocentric. However the structure at the centromere of the rearrangements could not be distinguish the origine of centromeric composition in single color FISH study.

In the literature, Lemyre E, et al (13) suggested that

most Robertsonian translocations are dicentric and also the location of chromosomal breaks leading to their formation occur in the acrocentric short arm. Furthermore, number of authors also confirm that in situ hybridization methods were used to clarify that the majority of Robertsonian translocations retain the alpha satellite DNA from both chromosomes involved and thus are structurally dicentric, with the breakpoints usually located in the acrocentric short arms, proximal to the NOR (3,7,9,14,15,16). These kind of dicentric chromosomes are stable during the cell division due to one of centromer inactivation (17). However, this translocation may result in unbalanced offspring in the next generation.

In our present study, some of case showed a distinct size of FISH signals, which may indicate the duplication or two different satellite DNA sequences in adjacent status. Therefore, these cases should be clarified for the satellite DNA composition of monocentric centromere sequence by using multicolor FISH study. These sequence composition may effect on the occurrence of interchromosomal effect raising chromosome aneuploidy and/or the behavior of translocated chromosome segregation in next generation.

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## REFERENCES

1. Therman E, Susman B, Denniston C. The nonrandom participation of human acrocentric chromosomes in Robertsonian translocations. *Ann Hum Genet* 1989;53: 49-65.
2. Frydman N, Romana S, Le Lorc'h M, Vekemans M, Frydman R, Tachdjian G. Assisting reproduction of infertile men carrying a Robertsonian translocation. *Hum Reprod* 2001;16: 2274-7.
3. Han J-Y, Choo KHA, Shaffer LG. Molecular cytogenetic characterization of 17 rob(13q14q) Robertsonian translocations by FISH, narrowing the region containing the breakpoints. *Am J Hum Genet* 1994;55: 960-7.
4. Pellestor F. Analysis of meiotic segregation in a man heterozygous for a 13;15 Robertsonian translocation and a review of the literature. *Hum Genet* 1990;85(1):49-54.
5. Craig J.M, Earnshaw W. C, and Vagnarelli P. Mammalian Centromeres: DNA Sequence, Protein Composition, and Role in Cell Cycle Progression *Experimental Cell Research* 1999;246: 249-62.
6. Choo K.H. Vissel B, Nagy A, Earle E, Kalitsis P *Nucleic Acids Res.* 1991;19, 1179-82.

7. Page SL, Shin JC, Han JY, Choo KH, Shaffer LG. Breakpoint diversity distinct mechanisms for Robertsonian translocation formation. *Hum Mol Genet* 1996;5(9):1279-88.
8. Gosden JR, Lawrie SS, Gosden CM. Satellite DNA sequences in the human acrocentric chromosomes: information from translocations and heteromorphisms. *Am J Hum Genet* 1981; 33:243-51.
9. Gravholt CH, Friedrich U, Caprani M, Jorgensen AL. Breakpoints in Robertsonian translocations are localized to satellite III DNA by fluorescence in situ hybridization. *Genomics* 1992; 14:924-30.
10. Waye JS, Willard HF. Human beta satellite DNA: genomic organization and sequence definition of a class of highly repetitive tandem DNA. *Proc Natl Acad Sci U S A* 1989;86(16):6250-4.
11. Kurnit DM, Roy S, Stewart GD, Schwedock J, Neve RL, Bruns GA, Van Keuren ML, Patterson D. The 724 family of DNA sequences is interspersed about the pericentromeric regions of human acrocentric chromosomes. *Cytogenet Cell Genet* 1986;43(1-2):109-16.
12. Worton RG, Sutherland J, Sylvester JE, Willard HF, Bodrug S, Dube I, Duff C, Kean V, Ray PN, Schmickel RD. Human ribosomal RNA genes: orientation of the tandem array and conservation of the 5' end. *Science* 1988; 239:64-8.
13. Lemyre E, der Kaloustian VM, Duncan AM. Stable non-Robertsonian dicentric chromosomes: four new cases and a review. *J Med Genet* 2001;38(1):76-9.
14. Wolff DJ, Schwartz S. Characterization of Robertsonian translocations by using fluorescence in situ hybridization. *Am J Hum Genet* 1992;50(1):174-81.
15. Sullivan BA, Jenkins LS, Karson EM, Leana-Cox J, Schwartz S. Evidence for structural heterogeneity from molecular cytogenetic analysis of dicentric Robertsonian translocation. *Am J Hum Genet* 1996;59(1):165-75.
16. Page SL, Shaffer LG. Nonhomologous Robertsonian translocations form predominantly during female meiosis. *Nature Genet* 1997; 15: 231-2.
17. Sullivan BA, Wolff DJ, Schwartz S. Analysis of centromeric activity in Robertsonian translocations: implications for a functional acrocentric hierarchy. *Chromosoma* 1994;103: 459-67.